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INITIAL STEPS ON PROCESSING A RENAL BIOPSY FOR ELECTRON MICROSCOPY & IMMUNOFLUORESCENCE

1. Obtain two kidney biopsy cores, if possible. Place the tissue in a petri dish or other appropriate cutting surface. Cut one renal tissue core into half and then cut two segments with a sharp razor blade from the middle section of the bisected tissue core (not from the ends).



- 2. Place one segment from the middle section into a vial of 2.5% glutaraldehyde in 0.1M NaCacodylate buffer for electron microscopy. Place the other cube from the middle section into a vial of Michel's Tissue Fixative for immunofluoresence microscopy.
- 3. Place all remaining tissue, i.e. pieces of the bisected core and entire other core, into a vial of 10% formalin for light microscopy.
- 4. Please contact the Renal Pathology and Electron Microscopy Laboratory at (203) 785-2750 to obtain addition Renal Biopsy Kits containing vials of formalin, glutaraldehyde in 0.1M NaCacodylate buffer and Michel's Fixative.
- 5. Label all vials with patient name, date and identifying numbers. Include the proper requisition sheet with each specimen. Vials with tissue do not need to be refrigerated during mailing, and should never be frozen.

Fixatives Caution: glutaraldehyde and formalin are poisonous, and vapors can be harmful. Extreme care should be taken with their use.